



Journal of Pharmaceutical Research

Research Article

Scientific Standardization of Various Extracts of *Chenopodium giganteum* D. LeavesShailin Dkhar^{1,*}, E Akila¹, V B Narayana Swamy¹, N Pruthvi¹¹Department of Pharmacognosy, R R College of Pharmacy, Chikkabanavara, Bangalore, 560090, Karnataka, India.

ARTICLE INFO

Article history:

Received 01.06.2022

Revised 23.06.2022

Accepted 29.06.2022

Published 22.09.2022

* Corresponding author.

Shailin Dkhar

shaidkhar@gmail.com[https://doi.org/](https://doi.org/10.18579/jopcr/v21i2.3)

10.18579/jopcr/v21i2.3

ABSTRACT

In the present research the Histological examination, physiological evaluation, phytochemical screening, total phenolic content and total flavonoid content and TLC of leaf extracts of *Chenopodium giganteum* has been studied. Ethanol, aqueous, chloroform, and pet ether extracts of *Chenopodium giganteum* was prepared. Phytochemical screening shows presence of carbohydrates, proteins, amino acids, glycosides like saponin and flavonoids, tannins, phenols, alkaloids, and steroids. The physiological evaluation shows total ash 15.12% w/w and acid insoluble ash value 7.46% w/w, loss of drying was found to be 8.1% w/w, extractive value of pet ether extract was found to be 2.3% w/w, chloroform extract 4.5% w/w, ethanol extract 7.2% w/w and water 10.2% w/w. The total flavonoid content of pet ether extract was found to be 17.2227 ± 0.0729 $\mu\text{g/ml}$, chloroform extract 23.7224 ± 0.0878 $\mu\text{g/ml}$, ethanol extract 49.8601 ± 0.0303 $\mu\text{g/ml}$ and water extract 64.7705 ± 0.0375 $\mu\text{g/ml}$. The total phenolic content of pet ether extract was found to be 14.477 ± 0.0226 $\mu\text{g/ml}$, chloroform extract 17.764 ± 0.0216 $\mu\text{g/ml}$, ethanol extract 19.518 ± 0.0173 $\mu\text{g/ml}$ and water extract 27.686 ± 0.0233 $\mu\text{g/ml}$. TLC of pet ether extract shows constituents having Rf values 0.548, 0.274, 0.1935, and 0.080, chloroform extract shows constituents having Rf values 0.765, 0.656, 0.468, 0.234, and 0.156, ethanol extract shows constituents having Rf values 0.815 and 0.584, and water extract shows constituents having Rf values 0.704 and 0.064.

Keywords: Histological examination; physiological evaluation; phytochemical screening; *Chenopodium giganteum*

INTRODUCTION

India is a great source of plant and animal richness Due to its diverse geographic and agro-climatic zones. It also boasts a diverse cultural legacy in addition to a diversified biosphere. Although the Indian health care system currently includes both conventional and alternative medicine, traditional medical practices like Ayurveda, Siddha, and Unani, as well as disorganized practices like folk medicine, have been thriving. Indian-based Ayurveda and Siddha make up around 60% of the nation's overall health care system and 75% of its rural residents.^{1,2} *Chenopodium giganteum*, a plant in the Amaranthaceae family, commonly referred to as tree spinach or labathua, is an annual, erect, many-branched shrub with an upper stem up to 5 cm diameter and a maximum height of 3 m. *Chenopodium amaranth tricolor* is another name for it.³⁻⁶ This plant, which grows at an elevation of 4,700 meters, has been used medicinally to treat a variety of illnesses linked to nutritional deficits. The plant,

which lowers the productivity of wheat, barley, mustard, and gramme crops, is a prevalent weed in waste areas and in fields throughout the summer and winter. It is also grown as a conventional leafy vegetable in India.⁷⁻⁹ Nearly no *Chenopodium giganteum* is grown for sale commercially. However, the reliable and substantial yield of *Chenopodium giganteum* suggests that it may be a plant of the future.¹⁰

Chenopodium giganteum young shoots and leaves may be prepared and eaten like spinach. The oxalic acid and saponins present in it are greatly destroyed by cooking if cooked for two minutes at 100 °C in boiling water.^{11,12} The leaves of *Chenopodium giganteum* can also be consumed raw in smaller amounts, like in a salad. Alternatively, the seeds can be crushed into flour and used with cereal flour to produce bread. The seeds can be prepared similarly to rice or quinoa. It also has decorative appeal because of its partly pink-colored leaves.¹³ It is also used in assessment for accumulation of heavy metals and to

investigate the usage of Fe, Zn, Cu, Ni, Cr, and Cd for phyto extraction of heavy metals.¹⁴ The plants belonging to the Amaranthaceae family and contains some saponins and oxalic acid, which, when present in excessive concentrations, can be harmful to human health. (e.g., Hemolysis or Kidney stone disease).¹⁵⁻¹⁸

The goal of this standardization is to guarantee the quality, safety, efficacy, and stability of final product by the careful, responsible selection and treatment of raw material. Additionally, there are no reports of comprehensive pharmacognostic research on leaves of this plant. This goal was considered when designing the current study, which aimed to evaluate the leaves scientifically. The investigation comprises physicochemical parameters, macroscopic and microscopic features, powder microscopic characteristics, TLC fingerprinting, and preliminary phytochemical screening. The data produced by this specific investigation provide pertinent pharmacognostical and physicochemical information required for accurate identification and verification of *Chenopodium giganteum* leaves.

MATERIAL AND METHODS

Collection, identification, and authentication of plant material

The leaves of *Chenopodium giganteum* (herbarium number XCH — 40416) plant was collected from the surrounding areas of Meghalaya. It was dried under shade and made into coarse powder. The plant material collected was identified and authenticated by Scientist (Dr) S. Mutheeswaran, M.Sc., M.Phil., Ph. D., Xavier Research Foundation, St Xavier's College, Tamil Nadu, India.

Morphological and macroscopic features

The fresh leaves of *Chenopodium giganteum* were examined for various macroscopic features like colour, odour and taste of leaves. Other external morphological characters like surface, base, margin, size, and shape of leaves were also studied. The microscopic examination of *Chenopodium giganteum* was done with the help of microscope. The air-dried plant material was then crushed into a coarse powder and used for further research work. The stomatal number and stomatal index of leaves of *Chenopodium giganteum* was also determined and evaluated by the methods referred from textbook authored by Kokate, Purohit and Gokhale.¹⁹

Physicochemical constants

The physical constants like ash value and extractive value helps in establishing the pharmacopoeial standards of the drug. The leaves of *Chenopodium giganteum* were examined for Physico-chemical constants like loss on drying, ash value and extractive value as per the methods mentioned in Pharmacopoeias.^{20,21}

Preparation of extracts

Successive Solvent extraction: Leaf of *Chenopodium giganteum* was dried and milled to a coarse powder. One kg of fresh plant material was grounded and defatted using petroleum ether. About 50g of the powdered air-dried defatted plant material was extracted subsequently with chloroform, pet ether, ethanol, and water in a Soxhlet apparatus. Each time before extracting with the next solvent, the marc was air dried below 50°C. The extracts were filtered, and the solvent obtained was evaporated at room temperature and accurate weight of the extracts was taken. The extractive value (%) was calculated with reference to air dried drug.

Preliminary Phytochemical screening

Preliminary phytochemical investigations of pet ether, chloroform, ethanol and water extracts were done to reveal the presence of different secondary metabolites like Proteins & Amino acids Carbohydrates, Steroids, Phenols, Saponins, Flavonoids, Alkaloids and Tannins.^{22,23}

Fluorescent analysis

Fluorescent analysis of both extracts and plant powders of *Chenopodium giganteum* leaves were carried out according to the method mentioned by Chase and Pratt et al. (1949) and Koshiet al. (1958) in day light and in UV light (254 nm and 365 nm). The plant powders and extracts were treated with different solvents and the fluorescence was observed in day light and in near and far UV light. About 10g of drug powder was taken in a petri dish and treated with different reagents and observed under different wavelengths i.e., ultraviolet and visible rays.²⁴

Total Phenol Content Determination

Using gallic acid as a standard, the total phenol concentration was calculated using the Folin-Ciocalteu test. In this process, 1.5 ml of Folin-reagent Ciocalteu's (FCR) diluted 1:10 v/v and 0.5 ml of plant extracts were combined. After 5 minutes, 1.5 ml of a solution containing 7% sodium carbonate was added. With distilled water, the final volume was made up to 10 ml, and they were then left to remain at room temperature for 90 minutes. A spectrophotometer was used to test the sample's absorbance at 750 nm in comparison to the blank. The entire experiment was done three times for accuracy and data expressed as mean standard deviation in terms of phenol content (gallic acid equivalent, or GAE) per g of dry weight.

Total Flavonoid Content Determination

Using quercetin as a standard, the total flavonoid content was calculated using the aluminium chloride technique. In this process, 4 ml of water and 1 ml of plant extracts

were combined. After 5 minutes, 0.3 ml of 10% Aluminum chloride, 0.3 ml of 5% Sodium nitrite and 1ml of 1 M Sodium hydroxide was added to the reaction mixture after the mixture had been incubated at room temperature for 6 minutes the final volume was made up to 10 ml with distilled water. A spectrophotometer was used to test the sample's absorbance at 510 nm in comparison to the blank. The entire experiment was done three times for accuracy, and data were expressed as mean standard deviation in terms of flavonoid content (Quercetin equivalent, or QE) per g of dry weight.²⁵

TLC of extracts

For TLC experiments, one gram of extracts was dissolved in methanol, filtered, and used. On aluminium plates that had already been coated with silica gel G, about 6µg of *Chenopodium giganteum* extract were applied. In a TLC chamber, the plate was produced using several solvent systems. Utilizing a photo documentation equipment, produced plates were seen and documented in both short and long UV.²⁶

RESULTS AND DISCUSSION

Morphology of *Chenopodium giganteum* leaves

The morphological data of *Chenopodium giganteum* showed the colour of young leaves was pink, magenta colour and older leaves were green with smooth under surface as shown in Figure 1. It has odour and bitter characteristic taste. The shape of leaves was extremely variable as simple, deltoid, ovate to lanceolate, upper entire, rhomboid, lower toothed or irregularly lobed. The size of leaf was about 1-3 cm; petioles were of 1-2 cm in length and as long as thick blade. Its length varied from 9 to 4.5 cm broad having dentate margin. It has acute apex and upto 5 cm base.

Microscopy of *C henopodium giganteum* leaves

The microscopic study of *Chenopodium giganteum* leaves was done with the help of microscope. The transverse section of the leaves reveals that the *Chenopodium giganteum* leaf is dorsiventral with palisade cells pointing upward toward the top epidermal layer. Nearly half of the leaflet is covered by the lengthy palisade cells. In the upper epidermis, there were spherical, thin-walled collenchymas in place of the thick-walled parenchymatous cells that were present there. The palisade layer is broken in the midrib area underneath the top epidermis by collenchymatous cells. Near the vascular bundles, thick-walled parenchymatous cells revealed the existence of spiral and annular vessels. While the cells in the lower epidermis are spherical, those in the upper epidermis are flattened. In the midrib area, behind the collenchyma cells, the vascular bundles are organized like a bunch of grapes. The transverse section and powder microscopy (magnification 10X) of leaves of *Chenopodium giganteum* is



Fig. 1: Morphological characters of leaves of *Chenopodium giganteum*

given below in Figure 2 and Figure 3.

Determination of leaf constants

The total stomatal number and stomatal index of upper epidermis of leaves was found to be 12 and 22.2% respectively and lower epidermis was found to be 11 and 12.5% respectively.

Determination of physicochemical constants

The total ash, acid-insoluble ash and water-soluble ash value and loss on drying of leaves of *Chenopodium giganteum* were evaluated and the results are mentioned in Table 1.

Table 1: Physico-chemical constants and leaf constants of leaves of *Chenopodium giganteum*

| Loss on drying | Total ash | Acid insoluble ash | Water soluble ash |
|----------------|-----------|--------------------|-------------------|
| 8.1% | 15.12% | 7.46% | 9.26% |

Extraction of plant material

The appearance and yield of different extracts of *Chenopodium giganteum* were evaluated and the results are

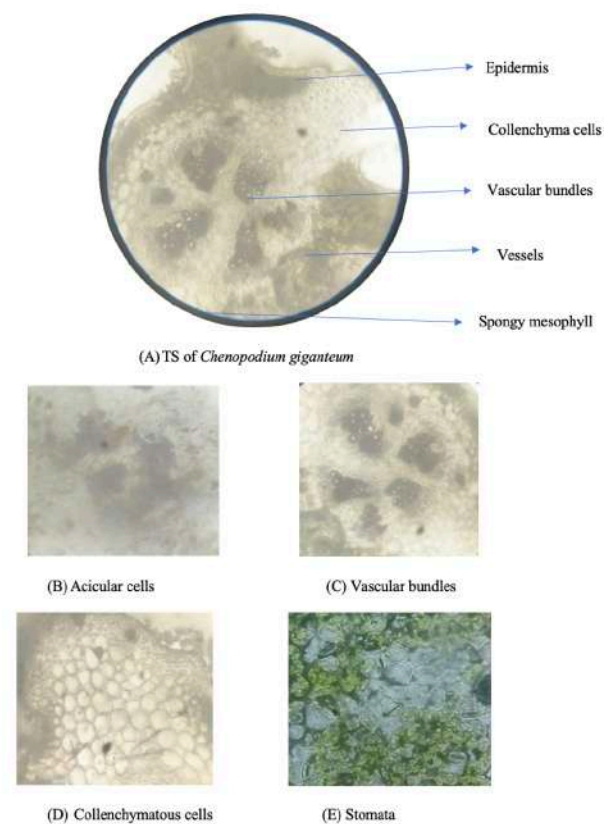


Fig. 2: A-E: Transverse section of leaves of *Chenopodium giganteum*

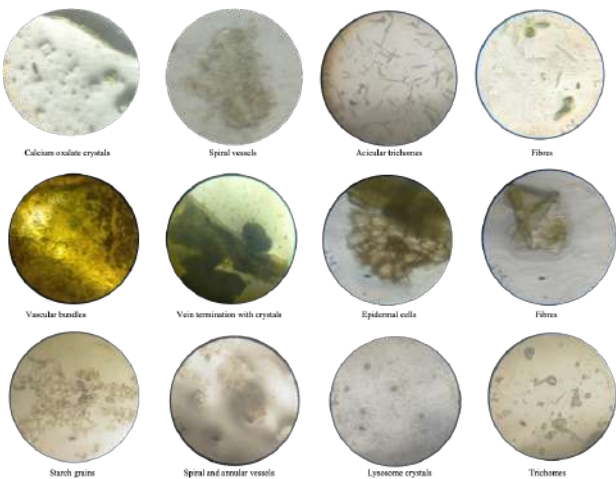


Fig. 3: Powder microscopy of leaves of *Chenopodium giganteum* (magnification 10X)

mentioned in Table 2.

Table 2: Yield of extracts obtained from successive extraction of leaves of *Chenopodium giganteum*

| Plant Name | Type of Extract | Appearance | State | Yield (% w/w) |
|-------------------------------------|-----------------|--------------------------------------------------------------------|-----------|---------------|
| <i>Chenopodium giganteum</i> leaves | Pet ether | Yellowish green | Semisolid | 2.3% w/w |
| | Chloroform | Greenish black | Semisolid | 4.5% w/w |
| | Ethanol | Dark green, black | Semisolid | 7.1% w/w |
| | Water | Dark brown black / Semi solid Dark Brown black / Dark brown, black | Semisolid | 10.2% w/w |

Preliminary phytochemical screening of extracts

Petroleum ether and chloroform extracts revealed the presence of steroids whereas ethanol and aqueous extracts indicated the presence of flavonoids, carbohydrates, saponins, proteins, alkaloids, phenols, steroids, and tannins respectively (Table 3).

Table 3: Preliminary phytochemical screening of leaf extracts of *Chenopodium giganteum*

| Chemical tests | <i>Chenopodium giganteum</i> leaf extracts | | | |
|-----------------------|--------------------------------------------|-------------|---------|-------|
| | Pet ether | Chloro-form | Ethanol | Water |
| Proteins & Amino acid | - | - | + | + |
| Carbohydrates | - | - | + | + |
| Steroids | + | + | - | - |
| Phenols | - | - | + | + |
| Saponins | - | - | + | + |
| Flavonoids | - | - | + | + |
| Alkaloids | - | - | + | + |
| Tannins | - | - | + | + |

‘-’ indicates absence and ‘+’ indicates presence

Fluorescent analysis

The selected plant is made into coarse powder and treated with required chemical reagents and observed under visible

and ultraviolet rays. The results are given in Table 4 and Table 5.

Table 4: Fluorescence analysis of powder of leaves of *Chenopodium giganteum*

| S.No. | Treatment | Day Light | Short UV (254nm) | LONG UV (366nm) |
|-------|-------------------------------------------|-----------------|------------------|-----------------|
| 1. | Powder | Green | Green | Green |
| 2. | Powder + Water | Green | Green | Dark green |
| 3. | Powder + 1NHCl | Green | Green | Dark green |
| 4. | Powder + 1NH ₂ SO ₄ | Green | Green | Dark green |
| 5. | Powder + 1NHNO ₃ | Green | Green | Dark-green |
| 6. | Powder + Acetic acid | Green | Green | Dark green |
| 7. | Powder + 1NNaOH | Green | Green | Dark green |
| 8. | Powder + 1NAlc.NaOH | Green | Green | Yellowish green |
| 9. | Powder + 1NKO ₂ H | Green | Green | Dark green |
| 10. | Powder + 1NAlc.KOH | Green | Green | Yellowish Green |
| 11. | Powder + Ammonia | Yellowish green | Green | Dark green |
| 12. | Powder + Iodine | Yellowish brown | Dark green | Dark green |
| 13. | Powder + FeCl ₃ | Yellowish brown | Dark green | Darkgreen |
| 14. | Powder + Ethanol | Green | Green | Yellowish green |

Table 5: Fluorescence analysis of leaf extracts of *Chenopodium giganteum*

| S.No | Extracts | Day Light | UV Light | |
|------|------------|----------------|-----------------|----------------|
| | | | Short 254nm | Long 365nm |
| 1 | Pet.ether | Green | Yellowish green | Yellowish |
| 2 | Chloroform | Greenish black | Darkgreen | Reddish brown |
| 3 | Ethanol | Greenish black | Greenishblack | Reddish brown |
| 4 | Water | Brownish dark | Green | Greenish black |

Total phenolic and Flavonoidal content

The total phenolic and flavonoidal content for aqueous, ethanol, chloroform, and petroleum ether extracts of *Chenopodium giganteum* were estimated and results are given in Table 6.

Table 6: Total phenolic content and total flavonoid content of leaf extracts of *Chenopodium giganteum*

| <i>Chenopodium giganteum</i> plant extract | Total phenolic content | Total flavonoid content |
|--------------------------------------------|------------------------|-------------------------|
| Pet ether | 14.477±0.0226 µg/ml | 17.2227±0.0729 µg/ml |
| Chloroform | 17.764±0.0216 µg/ml | 23.7224±0.0878 µg/ml |
| Ethanol | 19.518±0.0173 µg/ml | 49.8601±0.0303 µg/ml |
| Water | 27.686±0.0233 µg/ml | 64.7705±0.0375 µg/ml |

TLC studies of extracts

The extracts were undertaken for TLC profiling to assess the nature of phytochemicals present in it. A number of developing solvent systems were tried for all the extract and fractions. The solvent system, which gave the best resolution, was considered optimized, valid, and useful. The satisfactory resolution was obtained in the mobile phase mentioned in Table 7 and photo documentation is shown in Figure 4. The R_f values of different extracts was also calculated and are mentioned in Table 8. Blue and orange spots were observed which indicated the presence of phenolic compounds.²⁷

Table 7: Mobile phase for TLC studies of leaf extracts of *Chenopodium giganteum*

| Test extract | Solvent system | Number of Bands |
|--------------|------------------------------------------------------|-----------------|
| Pet ether | Toluene:Chloroform (1:1) | 05 |
| Chloroform | Toluene:Methanol (9:1) | 05 |
| Ethanol | Ethylacetate:Methanol:Glacialacetic acid (7:2.2:0.8) | 01 |
| Water | Ethyl acetate: Methanol: Water (5:1:2) | 02 |
| | Ethylacetate:Methanol:Glacialaceticacid (7:2.2:0.8) | 01 |
| | Ethyl acetate: Methanol: Water (5:1:2) | 02 |

Table 8: Rf values of different extracts of *Chenopodium giganteum*

| <i>Chenopodium giganteum</i> Leaf extracts | Color of the Spot | Rf value |
|--------------------------------------------|-------------------|----------|
| Pet ether | Light Blue | 0.548 |
| | Dark Blue | 0.352 |
| | Brown | 0.274 |
| | Brown | 0.1935 |
| | Blue | 0.080 |
| | Orange | 0.765 |
| Chloroform | Orange | 0.656 |
| | Blue | 0.468 |
| | Blue | 0.234 |
| | Purple | 0.156 |
| Ethanol | Orange | 0.815 |
| | Reddish orange | 0.584 |
| | Orange | 0.818 |
| | Light blue | 0.704 |
| Water | Dark blue | 0.064 |
| | Light blue | 0.731 |

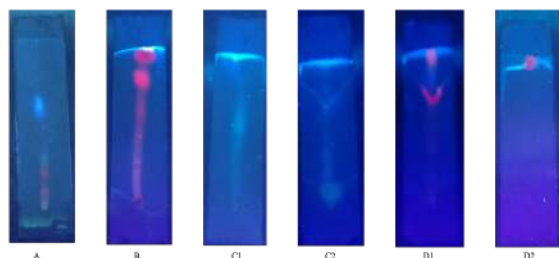


Fig. 4: TLC photo documentation of leaves of *Chenopodium giganteum*. (A) pet ether extract, (B) chloroform extract, (C1) water extract [Ethyl acetate:Methanol:Glacialaceticacid (7:2.2:0.8)], (C2) water extract [Ethyl acetate: Methanol: Water (5:1:2)], (D1) ethanol extract [Ethylacetate:Methanol:Glacialaceticacid(7:2.2:0.8)], (D2) ethanol extract [Ethyl acetate:Methanol: Water (5:1:2)]

CONCLUSION

In the current study, *Chenopodium giganteum* plant's leaves were examined for pharmacognostic characterisation, physiochemical parameter measurement, phytochemical screening, and TLC examinations of the crude extracts. The chosen plants were verified, and to prove their authenticity and purity, macroscopic examinations were carried out. To ascertain the fundamental cellular makeup of the leaf petiole, stem, kind of stomata, etc., a microscopic investigation was conducted. Physicochemical studies were carried out as per standard procedure such as ash value, acid insoluble ash values and extractive values. Fluorescence analysis was also carried out using different solvents. The important phyto-constituents were present as depicted in phytochem-

ical screening which are well-known for their medicinal potentials. The screening of leaves indicates the presence of high phenolic content and high flavonoid content which may be due to presence of phenol, flavonoid and tannin which possess antioxidant properties. Identification of the numerous phytochemical ingredients found in the raw medicine is aided by the TLC fingerprint profile. The TLC fingerprint profile aids in locating significant phyto-constituents as well. Thus, it is concluded that our study provides the data which helps to isolate, identify, and characterize the various medicinal potential of *Chenopodium giganteum* leaves.

REFERENCES

- Agarwal S. Clinically useful herbal drugs. 2005;p. 10–12.
- Sharma A, Shanker C, Tyagi LK, Singh M, Rao CV. Herbal medicine for market potential in India: an overview. *Acad J Plant Sci.* 2008;1(2):26–36. Available from: <http://www.idosi.org/ajps/1%282%2908/2.pdf>.
- Gelin Z, Sergei M, Steven EC. Chenopodiaceae (Flora of China). In: Zhengyi W, Raven PH, editors. *Chenopodiaceae*; vol. 5. Missouri Botanical Garden Press. 2003;p. 351–414.
- Yadav N, Vasudeva N, Singh S, Sharma SK. Medicinal properties of genus *Chenopodium* Linn. *Natural Product Radiance.* 2007;6(2):131–134. Available from: <http://nopr.niscpr.res.in/bitstream/123456789/7849/1/NPR%206%282%29%20131-134.pdf>.
- Pratap T, Joshi BD, Galvey NW. *Chenopods Chenopodium spp.* Promoting the conservation and use of underutilized and neglected crops. Rome, Italy. 1998.
- Rana TS, Narzary D, Ohri D. Genetic diversity, and relationships among some wild and cultivated species of *Chenopodium* L. (Amaranthaceae) using RAPD and DAMD methods. *Current Science.* 2010;98(6):840–846. Available from: <https://www.jstor.org/stable/24109856>.
- Panda H. *Handbook on Medicinal Herbs with Uses.* New Delhi. Asia Pacific Business Press. 2005.
- Pramila K, Neetu S, Anju R. Medicinal plants used in traditional health care system prevalent in Western Himalaya. *Indian J Traditional Knowle.* 2006;5(3):300–309.
- Patwardhan B, Vaidya M, Chorghade. Ayurveda and natural products drug discovery. *Current Science.* 2004;86(6):789–799. Available from: <https://www.jstor.org/stable/24109136>.
- Bhargava A, Shukla S, Ohri D. Evaluation of foliage yield and leaf quality traits in *Chenopodium* spp. *Euphytica.* 2007;153(1):199–213. Available from: <https://doi.org/10.1007/s10681-006-9255-8>.
- Ishii Y, Takiyama K. Characterization of oxalic acid in vegetables. *Analytical sciences.* 1991;7:811–815.
- Wang Z, Ando A, Takeuchi A, Ueda H. Effects of cooking conditions on the relationships among oxalate, nitrate, and lutein in spinach. *Food Science and Technology Research.* 2018;24(3):421–426. Available from: <https://doi.org/10.3136/fstr.24.421>.
- Hanelt P. *Mansfeld's World Database of Agricultural and Horticultural Crops.* 2017.
- Bhargava A, Shukla S, Srivastava J, Singh N, Ohri D. *Chenopodium: a prospective plant for phytoextraction.* *Acta Physiologiae Plantarum.* 2008;30(1):111–120. Available from: <https://doi.org/10.1007/s11738-007-0097-3>.
- Al-Saleh GF, El-Din AYG, Abbas JA, Saeed NA. Phytochemical and biological studies of medicinal plants in Bahrain: the family Chenopodiaceae-part 2. *International journal of pharmacognosy.* 1997;35(1):38–42. Available from: <https://doi.org/10.1076/phbi.35.1.38.13266>.
- Mroczek A. Phytochemistry and bioactivity of triterpene saponins from Amaranthaceae family. *Phytochemistry Reviews.* 2015;14(4):577–605. Available from: <https://doi.org/10.1007/s11101-015-9394-4>.

17. Siener R, Hönow R, Seidler A, Voss S, Hesse A. Oxalate contents of species of the Polygonaceae, Amaranthaceae and Chenopodiaceae families. *Food Chemistry*. 2006;98(2):220–224. Available from: <http://dx.doi.org/10.1016/j.foodchem.2005.05.059>.
18. Breuer D. Analytical Performance Issues: GESTIS Database: International Limit Values for Chemical Agents-A Readily Accessible Source of Occupational Exposure Limits (OELs). *Journal of Occupational and Environmental Hygiene*. 2010;7(7):37–42. Available from: <https://doi.org/10.1080/15459621003781231>.
19. Kokate CK, Purohit AP, Gokhale SB. Pharmacognosy. 2001.
20. The Ayurvedic Pharmacopoeia of India. Part II. 2008.
21. The Ayurveda Pharmacopoeia of India. Vol I. 2008.
22. Shaikh JR, Patil MK. Qualitative tests for preliminary phytochemical screening: An overview. *International Journal of Chemical Studies*. 2020;8(2):603–611. Available from: <https://doi.org/10.22271/chemi.2020.v8.i2i.8834>.
23. Khandelwal K. Practical pharmacognosy. 2008.
24. Pandey MK, Kumar A, Singh R, Tripathi M. Scientific standardization of leaves of *Chenopodium album* L. *Journal of Pharmacognosy and Phytochemistry*. 2016;5(5):1–6. Available from: <https://www.phytojournal.com/archives/2016/vol5issue5/PartA/5-4-6-331.pdf>.
25. Mathur R, Vijayvergia R. Determination of total flavonoid and phenol content in *Mimusops elengi* Linn. *International Journal of Pharmaceutical Sciences and Research*. 2017;8(12):5282–5285.
26. Arora SK, Itankar PR, Yende SR. Phytochemical screening and TLC studies of different extracts of *Chenopodium album*. *Journal of Ayurvedic and Herbal Medicine*. 2020;6(1):15–20. Available from: http://www.ayurvedjournal.com/JAHM_202061_05.pdf.
27. Sathya R, Kanaga N, Sankar P, Jeeva S. 2017.