



ORIGINAL ARTICLE

Development Studies On Microemulsion Gels Containing Herbal Excipients as Permeation Enhancers

Jimit Prakashkumar Shah¹, Subhabrata Ray^{1,*}

¹Department of Pharmaceutics, Krupanidhi College of Pharmacy, Bangalore, 560035, Karnataka, India

ARTICLE INFO

Article history:

Received 18.11.2019

Accepted 22.03.2020

Published 12.04.2020

* *Corresponding author.*

Subhabrata Ray

<https://doi.org/10.18579/jopcr/v19.1.jimith>

ABSTRACT

Objective: Acyclovir is an effective antiviral medication used in the management of Herpes Simplex Virus (HSV) infections. Although acyclovir sodium exhibits high solubility, it has low permeability, resulting in suboptimal bioavailability, necessitating elevated dosages, which can lead to increased side effects. The objective of this study was to formulate an oral microemulsion gel utilising a herbal excipient to enhance the permeability and bioavailability of acyclovir, thereby minimising the required doses and associated adverse effects.

Methods: Microemulsions were developed by systematically titrating different oil-to-surfactant ratios in an aqueous phase, as illustrated in a pseudo-ternary phase diagram. The evaluation included parameters, such as clarity, dilution shock, in vitro drug release, and ileal permeability. This method offers advantages such as reduced processing time, minimal exposure to high temperatures, and effective drug-loading efficiency.

Findings: The stable microemulsion gel composed of Captex 200, Tween 80, ethanol, water, propylene glycol, 100% Aloe vera, and 2% Carbopol demonstrated a fourfold enhancement in ileal permeability when compared to the acyclovir sodium solution.

Novelty: Aloe vera gel improves the permeation of microemulsions, thereby enhancing the intestinal permeability and bioavailability of BCS class III pharmaceuticals. This formulation has the potential to be administered through soft gel capsules.

Keywords: Acyclovir sodium; Aloe vera; BCS class III drugs; Permeability 200; Microemulsion gel; Permeability

INTRODUCTION

Microemulsions (MEs) are stable, fluid, transparent, or translucent homogenous dispersions with a quaternary composition of oil, aqueous phase, surfactant, and cosurfactant, forming a single optically isotropic dispersion with droplet diameters typically 10–100 nm^{1–5}. Microemulsions (MEs) have been extensively used to improve skin penetration of compounds with low water solubility and high lipophilicity¹. A surfactant/co-surfactant mixture was used to emulsify two immiscible phases, the oil and aqueous phases, resulting in a transparent and thermodynamically stable formulation⁶. Acyclovir [2-amino-9-(2-hydroxyethoxymethyl)-3H-purin-6-one], a synthetic purine nucleoside analog derived from guanine, is a BCS class III drug used to treat herpes simplex virus (HSV-1, HSV-2) and varicella-zoster virus⁷. Its pharmacokinetics after oral administration are highly variable, with peak plasma values of 0.46 to 0.83 µg/L

for a 200 mg dose or 0.63 to 1.21 µg/L for a 400 mg dose occurring 1.5 to 2.5 hours post-dose. Acyclovir is administered orally, via intravenous infusion, and topically⁸. This study developed an oral formulation of acyclovir using microemulsions and aloe vera gel as permeation enhancers, examining the impact of surfactants, cosurfactants, and a lipoidal system on the ileal permeability of acyclovir.

METHODOLOGY

Materials

Acyclovir in the form of sodium salt was generously gifted by Strides Arco Labs in India. Several oils were utilised, including Capmul MCM (Glyceryl Mono-Dicaprylate), Captex 200 (mixed diesters of caprylic/capric acids on propylene glycol) and Capmul PG-8 (Propylene glycol caprylate) were procured from ABITEC in the USA. The

surfactants incorporated in the formulation were Span 80 and Span 20 from Sigma-Aldrich in the USA, Tween 80 from Merck in India, Labrasol (polyglycolized glycerides) from Gattefosse in France, and Acconon CC-6 (ethoxylated caprylic and capric glycerol esters) from ABITEC in the USA. Cosurfactants included absolute ethanol sourced from Merck in India and Plurol Oleque (Polyglyceryl-6-dioleate) from Gattefosse in France. Aloe vera, used as a permeation enhancer, was gifted by Sagar Pharmaceuticals in India. Additionally, the polymer carbopol 974-P was sourced from Strides Arco Labs in India.

The study utilised a UV/VIS spectrophotometer, electronic balance, vortex shaker, micropipettes, and pH meter. Particle size and zeta potential analysers were used for particle analysis, and an IR Spectrometer was used for infrared spectroscopy. The water was semi-quartz distilled, and all chemicals and reagents were of A.R. grade, commercially procured, and used.

Physicochemical Study on the Drug

Melting point determination

Thiel's tube method of melting point determination in liquid paraffin was used in the present study⁹.

UV spectrum

UV spectrum scanning for the pure drug was conducted within the range of 200-300 nm. In the dilution medium of NaOH (0.1N), the maximum wavelength (λ_{max}) was observed at 265 nm. When using a dilution medium at 6.8 pH Phosphate buffer, λ_{max} was identified at 251.5 nm. Additionally, in a dilution medium of the Tyrode solution, the λ_{max} was determined to be 252.5 nm. Three standard calibration curves for acyclovir were plotted using 0.1N NaOH, phosphate buffer (pH 6.8), and Tyrode solution as the medium⁹.

Formulation

Selection of surfactant and co surfactant

A series of Surfactants with different HLB values have been screened such as Span 20, Span 80, Tween 80, Labrasol, Acconon CC-6 and Co- Surfactant like Ethanol and Plurol Oleque. Drug solubility studies were performed using different surfactants and co-surfactants. We found that Tween 80, in combination with a co-surfactant, ethanol, was the best for formulating a microemulsion.

Construction of Phase diagram

A pseudo-ternary phase diagram was created to study the formation of microemulsions using four components: oil surfactant, co-surfactant, and aqueous phase system. The system consisted of oil as Captex 200, surfactants such as Tween 80, co-surfactant ethanol, and double distilled water as the aqueous phase. A phase diagram was created

by drawing water dilution lines representing increasing water content and decreasing surfactant and co-surfactant levels. Water was titrated along the dilution lines from the surfactant apex to the opposite oil side of the triangle. If turbidity was followed by phase separation, the mixture was considered to be a biphasic dispersion. Clear and transparent mixtures were considered microemulsions. The transition point samples were marked as points in the phase diagram, and the area covered by these points was considered as the microemulsion region of existence^{10,11}.

Formulation Development

After the development of the phase diagram, different formulations were selected by keeping the total quantity of the formulation constant at 100% and varying all the components of the system. Each formulation was loaded with Acyclovir Na (5.1 mg/ml). All formulations were evaluated for different parameters such as clarity and dilution shock. The optimised formulation (which was selected from the data obtained from the evaluation studies) was further evaluated for in vitro release and intestinal permeability studies.

Effect of Aloe vera on Optimized Formulation

The optimised formulation was mixed with different concentrations of aloe vera (25, 35, 50, and 100% w/w). The formulations were then evaluated for in vitro release and intestinal permeability.

Effect of Aloe vera and Carbopol on Optimized Formulation

A microemulsion gel of the optimised formulation was formed with 2% carbopol 974-P which was further mixed with different concentrations of aloe vera (25%, 35%, 50%, and 100%). The formulations were then evaluated for in vitro release and intestinal permeability.

Evaluation Studies

Clarity

The clarity of all formulations was determined by measuring their absorbance and % transmittance at 650 and 400 nm, respectively⁸.

Dilution Shock test

Each formulation was diluted 10 and 25 times, and their absorbance and % transmittance were noted at 650 nm and 400 nm, respectively¹².

Particle size analysis

The particle sizes of the microemulsions were analysed using a Malvern particle size analyser.

Zeta potential

The zeta potential of the microemulsions was measured using a Malvern Zetasizer.

In vitro drug release study

The release study used a dialysis membrane (HIMEDIA 70 LA 393-10MT) activated with 5% EDTA solution and release medium. A phosphate buffer (pH 6.8) was used as the release medium. The membrane was suspended in a beaker containing 250 ml of buffer solution, stirred on a magnetic stirrer, and poured into a dialysis membrane tube. Samples were removed and diluted with phosphate buffer solution at intervals of 5, 10, 15, 30, 45, and 60 min and diluted to 10 ml with phosphate buffer solution. The absorbance of each sample was noted at 251.5 nm¹³

In vitro intestinal permeability study

The rats were euthanised with a chloroform overdose. The ileum from the small intestine was cut and removed and then placed in a Tyrode solution at 37 °C with aeration. The tissue was washed with Tyrode's solution to remove the extraneous luminal matter. A sample of 1.5 mg/ml was transferred to the tissue, tied, and placed in a beaker containing 50 ml of tyrode. The samples were removed at different intervals and diluted to 10 ml. The absorbance of these solutions was measured using a UV/VIS spectrophotometre at a wavelength of 252.5 nm, while the tyrode was kept blank. The same experiment was conducted for the blank to check for any absorbance in the tissue or tyrode solution¹⁴.

RESULTS AND DISCUSSION

Most drugs are structurally optimised during drug discovery studies to increase their solubility and efficiency of bio membrane permeation to reach their target receptors. While an increase in lipophilicity helps in better absorption, their delivery to the aqueous physiological milieu becomes difficult. On the other hand, some drugs have good water solubility but lack sufficient lipophilicity to effectively penetrate the lipoidal tissues, and absorption becomes rate-limiting for such candidates. Microemulsions, owing to their unique structure and dynamics¹⁵, afford efficient delivery of both lipophilic and hydrophilic drugs in aqueous and lipoidal environments. Owing to its absorption-enhancing effects, Aloe vera gel has been reported to effectively deliver poorly absorbable drugs through the oral route of drug administration¹⁶. This study investigated the potential of a synergy of microemulsion and aloe vera gel to improve the absorption of drugs with low solubility or permeability. Acyclovir, a synthetic purine nucleoside with low permeability, is a model drug that can be improved by formulating it in a microemulsion system and incorporating aloe vera gel. The formulation development process involves pre-formulation studies, analytical investigations, method selection, standardisation, validation, and preliminary trials.

The study also focused on selecting suitable oils, surfactants, and cosurfactants for oral administration, as well as studying their impact on the physicochemical and pharmaceutical properties of dosage forms.

The oral microemulsion of acyclovir underwent pre-formulation studies, including melting point and UV spectra analysis, to ensure it met Pharmacopoeial standards for melting point and wavelength of maximum absorption (λ_{max}). After ensuring the purity of the drug and verifying its λ_{max} in an appropriate solvent, a standard calibration curve was prepared, showing a linear relationship from 2 $\mu\text{g/ml}$ to 20 $\mu\text{g/ml}$ at 265 nm in 0.1N NaOH. Further analysis at 251.5 nm in phosphate buffer (pH 6.8 and 252.5 nm in Tyrode solution also showed linearity in the same concentration range. The study screened various surfactants and cosurfactants for drug solubility, with Tween 80 and ethanol being the most effective for formulating a microemulsion based on their HLB values and their combination with ethanol.

A pseudo-ternary phase diagram technique was used for microemulsion formulation. The oil was mixed with surfactants and cosurfactants and then titrated with water to form a turbid emulsion. The phase diagram was plotted as a triangle, with the weights of the oil, water, and surfactant mixture at different ratios. To optimise the formulation, specific points in the phase diagram were selected from the microemulsion zone. Each formulation was evaluated for clarity, dilution shock, in vitro release, and in vitro intestinal permeability. The results for each parameter are then discussed.

All three formulations exhibited clear, transparent, and pale yellow colours. Clarity is represented in terms of % transmission, with absorbance and % transmission noted at 650 nm and 400 nm, respectively. The best results were reported at 650 nm, with transmission exceeding 80% for all formulations. Visually, all formulations showed clear microemulsion formation owing to the minimal adjustment time required for surfactant and cosurfactant molecules to orient at the interface and achieve the necessary curvature for thermodynamic stability. The formulations were diluted with distilled water to 25X and 50X for dilution stock studies and their absorbance and % transmission was measured at 650 nm and 400 nm, respectively. All formulations maintained good clarity after dilution, with formulation 2 maintaining the best clarity even after 25X and 50X dilution (evidenced by a high %T near 90% and absorbance < 0.3). Additionally, formulation 2 had the best drug-loading capacity (5.1 mg) compared to the other formulations. The advantage of the system lies in its ability to maintain clarity and resist haziness after reaching the GIT, maintaining particle size in the microemulsion range, and increasing solubility and bioavailability due to the higher surface area of the nanodroplets (Table 1).

Table 1: Preformulation study on acyclovir sodium

Melting Point				
Reported			Observed	
256.5-257 °C			256 °C	
Drug solubility studies				
Sr. No	Category	Components	Drug loading capacity (mg)	
1	Surfactant	Labrasol	1.7	
		Acconon CC-6	2.1	
		Span 80	0.9	
		Span 20	1.1	
		Tween 80	3.6	
2	Co-Surfactant	Absolute Ethanol	2.8	
		Plurol Oleque	1.6	
		PEG-400	1.1	
		Glycerol	0.9	
3	Aqueous Solvent	Sorbitol	0.7	
		Water	97.5	
		Propylene glycol	10.7	
		Benzyl alcohol	2.3	
		Isopropyl alcohol	1.2	
4	Oil	Capmul MCM	0.7	
		Captex 200	1.4	
		CapmulPG-8	0.9	
Final Formulation Development Phase				
Formulation code	% oil	% S+C	% Aqueous Phase	Drug loading (mg)
FM 1	2.5	47.5	50	3.8
FM 2	3.5	46.5	50	5.1
FM 3	4.5	45.5	50	2.9
Clarity				
Formulation code	% Transmission at 650 nm		% Transmission at 400 nm	
FM 1	79.6		73.2	
FM 2	89.6		81.5	
FM 3	83.5		75.7	
Dilution shock test				
Formulation Code	Dilution 1:25 % Transmission		Dilution 1:50	
	AT 650nm	AT 400nm	AT 650nm	AT 400nm
FM 1	91.7	86.4	94.3	90.2
FM 2	93.5	90.7	96.7	92.3
FM 3	92.9	88.3	96.1	92.5
Formulation code	Dilution 1:25 Absorbance		Dilution 1:50	
	AT 650 nm	AT 400 nm	AT 650 nm	AT 400 nm
FM 1	0.038	0.064	0.025	0.044
FM 2	0.029	0.041	0.022	0.038
FM 3	0.031	0.054	0.023	0.038

The in vitro drug release study revealed that the drug release after 1 h for the optimised microemulsion formulation (F2) was $55\% \pm 0.952$, compared to $91\% \pm 1.938$ for the pure drug (Table 2).

This indicates a slowdown in drug release from the microemulsion, likely due to reduced diffusion of the drug's sodium salt through the lamellar surfactant/cosurfactant layers, which increases the microenvironment viscosity and may suggest migration of the drug into the oily phase or entrapment in the surfactant-cosurfactant layers. When different concentrations of aloe vera (ranging from 25% to 100%) were mixed with the optimised formulation, drug release increased with higher aloe vera concentrations: $62\% \pm 1.42$ for 25% aloe vera, $69\% \pm 1.67$ for 35% aloe vera, $73\% \pm 6.37$ for 50% aloe vera, and $80\% \pm 9.66$ for 100% aloe vera (Table 2). This increase is likely due to the hydrophilic Aloe vera causing relaxation of the lamellar structure of the surfactant-cosurfactant layers, allowing more drug to diffuse out. However, in the presence of 2% carbopol and varying concentrations of aloe vera, there was a decrease in drug release: $65\% \pm 14.03$ with 25% aloe vera, $61\% \pm 11.02$ with 35% aloe vera, $52\% \pm 7.83$ with 50% aloe vera, and $45\% \pm 6.74$ with 100% aloe vera (Table 3).

This reduction was attributed to the higher viscosity of 2% carbopol, which lowered the diffusion coefficient. After 1 h of permeation, most of the microemulsion formulations showed enhanced permeation compared to the drug solution. Permeation after 1 h from the optimised formulation was $31\% \pm 0.91$, as opposed to only $18\% \pm 1.09$ for the pure drug (Table 2). This is in line with the BCS classification of the drug into class III (high solubility, 91% drug release after 1 h; low permeability). Different concentrations of aloe vera gel, ranging from 25% to 100%, were mixed with the optimised formulation. Drug permeation after 1 h from 25% aloe vera was $48\% \pm 1.27$, from 35% aloe vera was $55\% \pm 1.31$ (Table 2), from 50% aloe vera was $59\% \pm 1.05$, and from 100% aloe vera was $76\% \pm 4.43$ (Table 3). Aloe vera enhances drug permeation by reversibly opening tight junctions, allowing better drug penetration. As aloe vera concentration increased, drug permeation also increased. However, its impact on ileal membrane structure remains unexplored. Drug permeation from microemulsions with 2% carbopol and different aloe vera concentrations showed a $29\% \pm 1.89$ rate at the end of 1 h, with 35% aloe vera and 2% carbopol $35\% \pm 1.13$, 50% aloe vera and 2% carbopol $44\% \pm 1.11$, and 100% aloe vera and 2% carbopol $58\% \pm 9.82$ (Table 3). Carbopol caused a 20% reduction in permeability, affecting drug release and potentially reducing drug availability owing to increased viscosity.

CONCLUSION

Acyclovir was formulated into a microemulsion gel system, which was then transformed into a microemulsion gel with

Table 2: In vitro drug release and permeation

In vitro drug release for pure drug				
Time (min)	% CDR Average (Pure drug in distilled water at 5.1 mg/mL)		% CDR Average (Optimised formulation (f2) in Phosphate buffer)	
5 min	20.366 ± 6.215		12.43 ± 1.030	
10 min	30.516 ± 2.823		14.29 ± 1.411	
15 min	39.34 ± 6.161		18.75 ± 0.610	
30 min	54.43 ± 5.837		33.3 ± 1.170	
45 min	72.653 ± 6.082		44.4 ± 1.056	
60 min	91.416 ± 1.938		55.36 ± 0.952	
In vitro drug permeation for pure drug				
Time in min	% CDR Average (Pure drug in distilled water at 5.1 mg/mL)		% CDR Average (Optimized formulation f2-in tyrode solution)	
5 min	6.77 ± 0.412		8.70 ± 0.496	
10 min	8.6 ± 0.512		13.42 ± 1.276	
15 min	12.33 ± 1.597		16.98 ± 0.25	
30 min	13.52 ± 1.266		20.66 ± 0.502	
45 min	15.46 ± 0.705		24.65 ± 1.524	
60 min	18.26 ± 1.097		31.3 ± 0.901	
In vitro drug release and permeation for formulation				
Time (min)	Drug release % CDR (Formulation containing drug solution in distilled water at 5.1 mg/ml and 100% aloe vera)	Drug permeation % CDR (Formulation containing drug solution in distilled water at 5.1 mg/ml and 100% aloe vera)		
5	43.6	11.2		
10	54	16		
15	66.4	20.8		
30	78.8	30.4		
45	87.2	± 38		
60	95.6	9 ± 40.4		
Release study of optimized formulation				
Time in min	% CDR Average (25% Aloe vera)	% CDR Average (35% Aloe vera)	% CDR Average (50% Aloe vera)	% CDR Average (100% Aloe vera)
5 min	13.87 ± 1.330	20.57 ± 1.866	21.93 ± 0.230	20.67 ± 3.422
10 min	19.82 ± 0.346	26.17 ± 0.739	28.1 ± 1.11	27.84 ± 3.424
15 min	28.63 ± 0.461	31.70 ± 0.950	38.82 ± 3.011	41.07 ± 4.411
30 min	42.78 ± 0.956	47.68 ± 0.493	53.99 ± 3.677	55.79 ± 1.200
45 min	52.19 ± 0.661	60.09 ± 2.201	66.71 ± 4.232	73.29 ± 9.031y
60 min	61.86 ± 1.428	69.07 ± 1.679	73.56 ± 6.372	79.78 ± 9.664

Carbopol and Aloe vera gel. The microemulsion showed improved drug release, with Aloe vera gel enhancing permeation. The study concluded that Aloe vera gel can significantly improve the intestinal permeability and bioavailability of BCS class III drugs through a microemulsion platform. Future studies could include whole-animal pharmacokinetic studies and human clinical trials.

Table 3: Permeation, release and permeability study of optimized formulation

Permeation study of optimized formulation									
Time in min	%CDR Average (25% Alora)	(25% Alora)	%CDR Average (35% Alora)	(35% Alora)	%CDR Average (50% Alora)	(50% Alora)	%CDR Average (100% Alora)	(100% Alora)	
5	11.38 ± 0.792		12.16 ± 2.772		14.66 ± 2.552		16.36 ± 2.502		
10	14.41 ± 0.547		14.97 ± 1.390		17.59 ± 0.760		21.76 ± 2.244		
15	17.38 ± 0.665		19.76 ± 0.465		22.17 ± 0.918		26.11 ± 1.654		
30	24.94 ± 1.530		27.54 ± 1.640		28.43 ± 1.029		30.17 ± 1.031		
45	37.80 ± 1.096		39.28 ± 0.906		42.08 ± 1.974		48.68 ± 1.244		
60	48.67 ± 1.270		54.80 ± 1.315		59.25 ± 1.057		76.39 ± 4.432		
Release study of optimized formulation with 2% carbopol and 25% aloe vera									
Time in min	%CDR Average (2% Carbopol and 25% Aloe vera)	(2% Carbopol and 25% Aloe vera)	%CDR Average (2% Carbopol and 35% Aloe vera)	(2% Carbopol and 35% Aloe vera)	%CDR Average (2% Carbopol and 50% Aloe vera)	(2% Carbopol and 50% Aloe vera)	%CDR Average (2% Carbopol and 100% Aloe vera)	(2% Carbopol and 100% Aloe vera)	
5 min	7.20 ± 0.987		4.75 ± 0.447		6.38 ± 0.626		4.94 ± 1.939		
10 min	16.60 ± 1.612		12.71 ± 0.86		10.86 ± 0.419		8.23 ± 2.574		
15 min	22.67 ± 3.026		22.29 ± 1.124		20.45 ± 0.929		19.72 ± 5.015		
30 min	33.7 ± 2.683		29.79 ± 1.291		27.73 ± 1.136		26.22 ± 5.675		
45 min	49.50 ± 2.058		43.81 ± 1.510		39.97 ± 3.310		38.36 ± 7.379		
60 min	65.83 ± 14.035		61.47 ± 11.021		52.24 ± 7.833		45.21 ± 6.740		
Permeability study of optimized formulation									
Time in min	%CDR Average (2% Carbopol and 25% Aloe vera)	(2% Carbopol and 25% Aloe vera)	%CDR Average (2% Carbopol and 35% Aloe vera)	(2% Carbopol and 35% Aloe vera)	%CDR Average (2% Carbopol and 50% Aloe vera)	(2% Carbopol and 50% Aloe vera)	%CDR Average (2% Carbopol and 100% Aloe vera)	(2% Carbopol and 100% Aloe vera)	
5 min	1.66 ± 0.650		3.05 ± 0.783		6.11 ± 0.57		11.55 ± 2.785		
10 min	4.09 ± 0.525		5.43 ± 0.981		8.75 ± 0.441		18.82 ± 4.421		
15 min	6.84 ± 1.086		7.82 ± 0.842		12.29 ± 1.006		24.03 ± 5.069		
30 min	13.69 ± 1.615		17.42 ± 1.011		22.49 ± 0.903		33.95 ± 6.262		
45 min	20.08 ± 1.551		24.50 ± 0.953		31.34 ± 0.794		43.92 ± 8.129		
60 min	28.89 ± 1.891		35.43 ± 1.131		44.73 ± 1.119		57.87 ± 9.825		

REFERENCES

- Lawrence MJ, Rees GD. Microemulsion-based media as novel drug delivery systems. *Advanced drug delivery reviews*. 2000;45(1):89–121. Available from: [https://doi.org/10.1016/S0169-409X\(00\)00103-4](https://doi.org/10.1016/S0169-409X(00)00103-4).
- Tenjarla S. Microemulsions: an overview and pharmaceutical applications. *Crit Rev Ther Drug Carrier Syst*. 1999;16:461–521. Available from: <https://pubmed.ncbi.nlm.nih.gov/10635455/>.
- Eccleston GM. Emulsions and Microemulsions. In: and others, editor. *Encyclopedia of Pharmaceutical Science and Technology*; vol. 2. CRC Press. 2007;p. 1066–85. Available from: <https://www.taylorfrancis.com/chapters/edit/10.1081/E-EPT4-120050265/emulsions-microemulsions-gillian-eccleston>.
- Lawrence MJ, Warisnoicharoen W. 2006.
- Coneac G, Vlaia V, Olariu I, Mut AM, Anghel DF, Ilie C, et al. Development and evaluation of new microemulsion-based hydrogel formulations for topical delivery of fluconazole. *American Association of Pharmaceutical Scientists*. 2015;16:889–904. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4508307/pdf/12249_2014_Article_275.pdf.
- Rangsimawong W, Wattanasri P, Tonglairoom P, Akkaramongkolporn P, Rojanarata T, Ngawhirunpat T, et al. Development of Microemulsions and Microemulgels for Enhancing Transdermal Delivery of Kaempferia parviflora Extract. *AAPS PharmSciTech*. 2018;19:2058–2067. Available from: <https://pubmed.ncbi.nlm.nih.gov/29687193/>.
- Kurokawa M, Nagasaka K, Hirabayashi T, Uyama SI, Sato H, Kageyama T, et al. Efficacy of traditional herbal medicines in combination with acyclovir against herpes simplex virus type 1 infection in vitro and in vivo. *Antiviral research*. 1995;27(1-2):19–37. Available from: <https://pubmed.ncbi.nlm.nih.gov/7486956/>.
- Ghosh PK, Majithiya RJ, Umrethia ML, Murthy RS. Design and development of microemulsion drug delivery system of acyclovir for improvement of oral bioavailability. *AAPS PharmSciTech*. 2006;7:172–179. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2750519/pdf/12249_2008_Article_730172.pdf.
- phannacoepia US. Technetium-99m-disofenin injection. . Rockville, MD. United States Pharmacopeia Convention, USP-NF. 1995. Available from: https://doi.usp.org/USPNF/USPNF_M80670_04_01.html.
- Subramanian N, Ray S, Ghosal SK, Bhadra R, Moulik SP. Formulation design of Self-micro emulsifying drug delivery system for improved oral bioavailability of Celecoxib. *Biological and Pharmaceutical Bulletin*. 2004;27(12):1993–1999. Available from: https://www.jstage.jst.go.jp/article/bpb/27/12/27_12_1993/_article.
- Lewis GA, Mathieu D, Phan-Tan-Luu R. Pharmaceutical Experimental Design. 1st ed. and others, editor; Boca Raton. CRC press. 1998. Available from: <https://www.taylorfrancis.com/books/mono/10.1201/9780203508688/pharmaceutical-experimental-design-gareth-lewis-didier-mathieu-roger-phan-tan-luu>.
- Li P, Ghosh A, Wagner RF, Krill S, Joshi YM, Serajuddin AT. Effect of combined use of nonionic surfactant on formation of oil-

- in-water microemulsions. *International journal of pharmaceutics*. 2005;288(1):27–34. Available from: <https://www.sciencedirect.com/science/article/abs/pii/S0378517304005356>.
13. Gibson M. Pharmaceutical preformulation and formulation. vol. 199 of *Drugs and the Pharmaceutical Sciences*. 2001;p. 1–560. Available from: <https://www.routledge.com/Pharmaceutical-Preformulation-and-Formulation-A-Practical-Guide-from-Candidate-Drug-Selection-to-Commercial-Dosage-Form/Gibson/p/book/9781420073171?srsltid=AfmBOorEJ0MvzhQp0OjVbq0ullnlhpUJmfjSXc06PCxVbpXQVEpyXNq3>.
 14. Welling PG, Tse FL. *Pharmacokinetics: regulatory, industrial, academic perspectives*. Marcel Dekker Inc. 1988. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1365164/pdf/brjclinpharm00003-0123.pdf>.
 15. Ruckenstein E. Thermodynamic approaches to microemulsions. *Journal of colloid and interface science*. 1998;204(1):143–150. Available from: <https://www.sciencedirect.com/science/article/abs/pii/S0021979798955552>.
 16. Hamman JH. Composition and applications of Aloe vera leaf gel. *Molecules*. 2008;13(8):1599–1616. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6245421/pdf/molecules-13-01599.pdf>.